

Cytogenetic studies of the F₁ hybrids of *Capsicum annuum* with *C. chinense* and *C. baccatum*

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Summary. Partially sterile interspecific hybrids were obtained between *C. annuum* var. ‘cerasiformis’ and *C. chinense* var. ‘mishme’ (H₁), and *C. annuum* var. ‘cerasiformis’ and *C. baccatum* var. ‘pendulum’ (H₂). Morphologically the F₁ hybrids were intermediate between the corresponding parents. Meiosis was irregular in the two F₁ hybrids. Cytological analysis of the two F₁ hybrids revealed that the genome of *C. annuum* differs from *C. chinense* by two translocations and some minor structural alterations and from *C. baccatum* by two translocations, a single inversion and some minor structural alterations. Isolation barriers such as hybrid inviability, weakness and hybrid breakdown in the H₁ hybrid and, in addition, desynapsis in the H₂, were operative in these taxa. The differences between the present findings and those reported earlier on the two F₁ hybrids were attributed to differences in the genetic architecture of the taxa employed in hybridization.

Key words: *Capsicum annuum*, *C. chinense*, *C. baccatum* – Interspecific hybrids – Chromosome pairing – Genome homologies

Introduction

Chili pepper (*Capsicum* L.), an important condiment, is more or less cosmopolitan in distribution and is cultivated as a cash crop not only in India but in many parts of the world.

In spite of its economic importance, the cytogenetic work necessary for a proper understanding of the interspecific relationships in the genus has not been carried out. Though crossability relationships among some taxa of this genus have been reported (Smith and Heiser 1951, 1957; Hirose et al.

1960; Lippert et al. 1966; Eshbaugh, 1964, 1975; Somanatha Pillai et al. 1977; Yamamoto 1978), these were mostly confined to the breeding behaviour of the F₁ hybrids. Nevertheless, information pertaining to the cytological analysis of a few interspecific hybrids including the F₁ hybrids of *C. annuum* × *C. chinense* and *C. annuum* × *C. baccatum* is available (Ohta 1961; Venkateswara Rao and Yesoda Raj 1974; Pickersgill 1971, 1977; Koompai 1976; Tanksley 1984; Egawaa and Tanaka 1984, 1986; Aniel Kumar 1984; Panda 1985).

In the present paper we describe cytological observations on F₁ hybrids of *C. annuum* × *C. chinense* and *C. annuum* × *C. baccatum*, and relate these to some of the differences previously recorded on these two F₁ hybrids (Pickersgill 1971, 1977; Koompai 1976; Tanksley 1984; Egawa and Tanaka 1984, 1986).

Materials and methods

Seeds of *C. annuum* var. ‘cerasiformis’ were obtained from N. S. R. Murthy, Agricultural Research Station, Lam Farm, Guntur, India. Seeds of *C. chinense* var. ‘mishme’ and *C. baccatum* var. ‘pendulum’ were provided by E. Pochard, Montfavet, Avignon, France. The three species were selfed and maintained for several generations before the hybridization programme was began. Reciprocal crosses were attempted between *C. annuum* and *C. chinense* and *C. annuum* and *C. baccatum* respectively. Two F₁ interspecific hybrids, *C. annuum* × *C. chinense* and *C. annuum* × *C. baccatum*, were obtained and these provided the material for the present study. F₁ hybrids of *C. annuum* var. ‘cerasiformis’ × *C. chinense* var. ‘mishme’ and *C. annuum* var. ‘cerasiformis’ × *C. baccatum* var. ‘pendulum’ are referred to as H₁ and H₂, respectively, in the text.

For cytological analysis, young flower buds from both the parents and the F₁ hybrids were fixed in a 1:3 acetic acid and alcohol mixture. After 24 h, they were transferred to 70% alcohol and stored in a refrigerator. Smears were made in 2% acetocarmine to study meiosis. Pollen fertility – sterility was determined by staining mature and ripe anthers with 2% acetocarmine. Well-filled and stained grains were scored as fertile while shrunken, unstained or half-stained grains were considered to be sterile.

Results

Crossability

Though reciprocal crosses between *C. annuum* and *C. chinense*, and *C. annuum* and *C. baccatum* yielded seeds, viable F₁ hybrids could be raised only from the crosses *C. annuum* × *C. chinense* (H₁) and *C. annuum* × *C. baccatum* (H₂). The seeds from the reciprocal crosses either did not germinate or those few that did germinate died after a few days (Table 1).

Morphology of parents and hybrids

C. annuum, *C. chinense*, and *C. baccatum* conform to the taxonomic description of Eshbaugh (1980) and Anonymous (1983). The two F₁ hybrids (H₁ and H₂) were rather weak and morphologically they were intermediate between the corresponding parents. In a few characters they resembled either one of the other of the parents.

Cytology of parents and hybrids

Chromosome pairing and behaviour was studied in 300 PMCs, at both diakinesis and metaphase I, in all

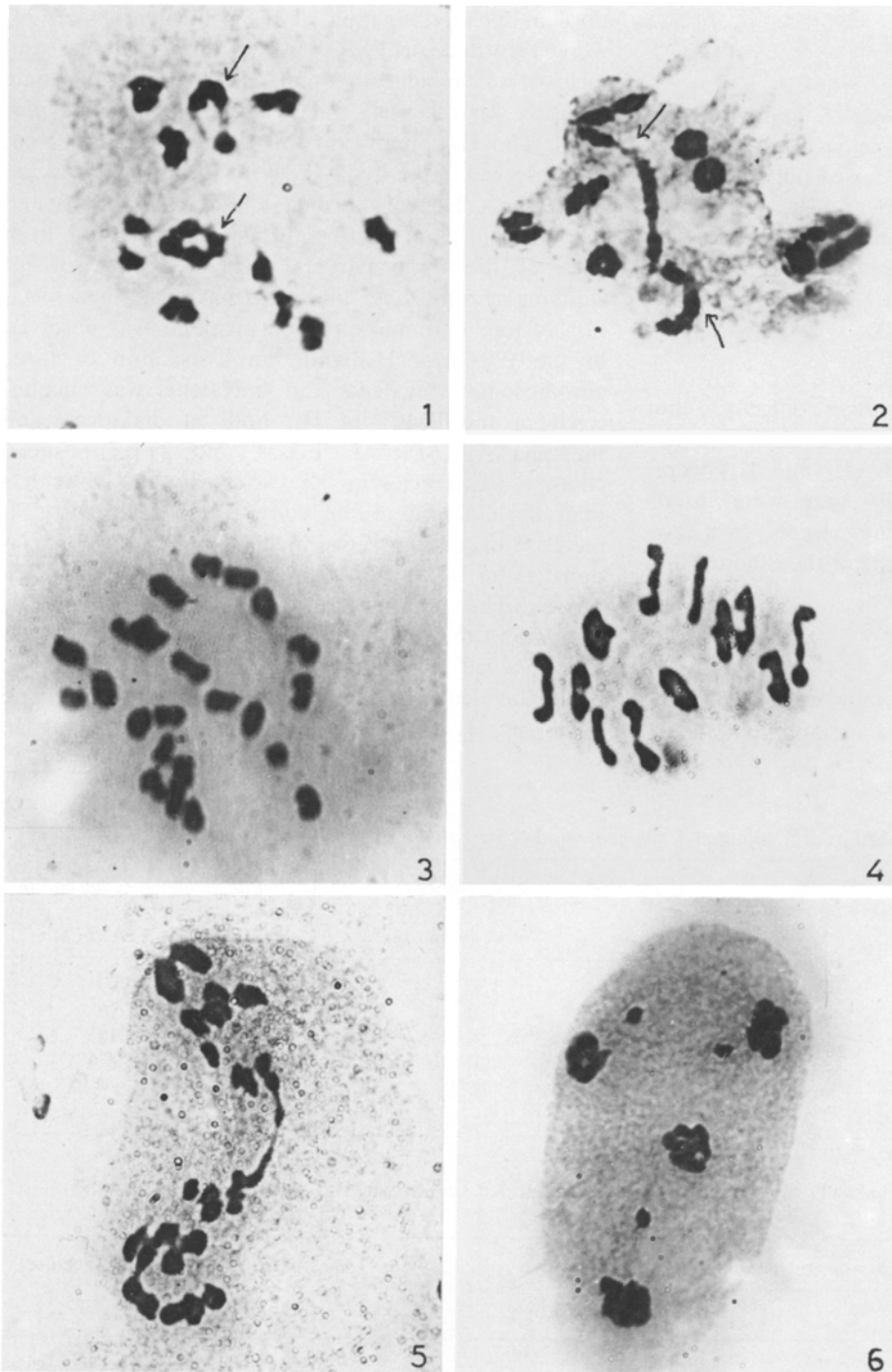
taxa under investigation. Twelve bivalents (2n=24) were recorded in PMCs in all three parents, and meiosis was regular in them. However, chromosome synapsis was relatively poor and meiosis was irregular in H₁ and H₂. Significant interplant differences were not observed in the two F₁ hybrids (H₁ and H₂), non in the parents. Hence, the data for three H₁ plants and five H₂ plants, and for 10 plants chosen at random from each of the three parental taxa, were pooled for studying chromosome pairing behaviour. An association of four chromosomes and bivalents was observed in the PMCs of H₁ while an association of three chromosomes, bivalents and univalents was encountered in the PMCs of H₂, both at diakinesis and metaphase I (Table 2, Figs. 1 and 2). The mean chiasma frequencies in the two F₁ hybrids were less than those found in the corresponding parents (Table 2). A maximum of two higher chromosome associations (IV or III) per PMC was observed in H₁ and H₂. These higher chromosome associations were mostly chains and the nucleolus organising chromosome was not involved in the interchange. The occurrence of twelve bivalents in a PMC was higher in H₁ (37%) and lower in H₂ (21%) (Fig. 4). Univalents ranging in

Table 1. Results on crossability relationships of *C. annuum*, *C. chinense* and *C. baccatum*

Particulars	<i>C. annuum</i>	<i>C. chinense</i>	<i>C. annuum</i>	<i>C. baccatum</i>
	× <i>C. chinense</i>	× <i>C. annuum</i>	× <i>C. baccatum</i>	× <i>C. annuum</i>
No. of pollinations made	200	150	200	250
No. of fruits obtained	20	10	20	16
Average no. of seeds/fruit	20	16	13	18
No. of seeds germinated	35	10	40	12
Percentage of seed germination	8.75	6.25	15.38	4.17
No. of hybrid plants surviving	3	—	5	—

Table 2. Mean chromosome pairing, chiasma frequency at metaphase I, pollen stainability and seed set in parents and their F₁ hybrids

Parents and hybrids	Chromosome associations				Chiasmata	% of pollen stainability	Seed set
	IV	III	II	I			
<i>C. annuum</i> var. 'cerasiformis'	—	—	12.00	—	17.36 ± 0.19 (15–19)	83.70 ± 0.42 (76–86)	46 ± 0.15 (26–52)
<i>C. chinense</i> var. 'mishme'	—	—	12.00	—	19.64 ± 0.20 (17–24)	92.70 ± 0.18 (86–94)	35 ± 0.14 (20–45)
<i>C. baccatum</i> var. 'pendulum'	—	—	12.00	—	19.32 ± 0.24 (17–22)	89.80 ± 0.28 (79–91)	40 ± 0.20 (18–45)
<i>C. annuum</i> × <i>C. chinense</i>	0.63 ± 0.03 (0–2)	—	10.73 ± 0.06 (8–12)	—	16.99 ± 0.02 (9–24)	42.00 ± 2.05 (30–49)	20 ± 0.10 (6–28)
<i>C. annuum</i> var. 'cerasiformis' × <i>C. baccatum</i> var. 'pendulum'	—	0.42 ± 0.05 (0–2)	7.88 ± 0.20 (5–12)	6.98 ± 0.43 (0–16)	11.57 ± 0.54 (6–20)	42.60 ± 0.62 (26–49)	13 ± 0.25 (4–15)



Figs. 1-6. Meiosis in interspecific hybrids of *Capsicum*. **1** Diakinesis showing 2 IV (t) + 8 II (H₁) ($\times 1,600$); **2** Diakinesis showing 2 III (t) + 9 II (H₂) ($\times 1,600$); **3** Metaphase I showing 2 II + 20 I (H₂) ($\times 1,600$); **4** Metaphase I showing 12 II (H₁) ($\times 1,600$); **5** Anaphase I showing bridge (H₁) ($\times 1,600$); **6** Telophase II showing laggards (H₂) ($\times 1,600$)

numbers from 0-24 were recorded in H₂ and these were scattered throughout the cytoplasm (Fig. 3).

At anaphase I 200 PMCs were scored for chromosome disjunction for both F₁ hybrids. Irregular chromosome disjunction and laggards were observed in 60% of the H₁ PMCs and a single persistent bridge (Fig. 5) and fragments (0-2) in addition to irregular chromosome segregation and laggards were recorded in

75% of the H₂ cells. The irregularities persisted until the end of the second meiotic division though their frequencies were lower than those found in anaphase I (Fig. 6). Micronuclei varying in numbers from 0-4 were also observed in some of the PMCs of the two F₁ hybrids.

Pollen fertility and seed set was very low in the two F₁ hybrids compared to the corresponding parents

(Table 2). The selfed seeds of the two hybrids were shrivelled with a depression at the centre, consequently these did not germinate. A few seeds were obtained when the H_2 was backcrossed to the *C. annuum* parent and though some of them did germinate these did not survive until maturity. On the other hand, the *C. annuum* × *C. chinense* hybrid (H_1) when backcrossed to either of the parents, and H_2 when backcrossed to the *C. baccatum* parent, did not set seed.

Discussion

For assessing interspecific relationships reciprocal crosses were made between both *C. annuum* and *C. chinense*, and *C. annuum* and *C. baccatum*. Viable F_1 hybrids were obtained only when *C. annuum* was the seed parent in each case. However, a few seeds were obtained in the reciprocal crosses for either of the two combinations and some of them did germinate. Nevertheless, these died after a few days. F_2 progeny could not be raised from either of the two hybrids and the few backcross seedlings obtained from H_2 did not survive until maturity. All this suggests the manifestation of hybrid inviability and weakness in the prezygotic stage preventing hybrid formation in the reciprocal direction and the operation of hybrid sterility and hybrid breakdown in the post-zygotic stage of hybrid development resulting in the failure to obtain F_2 and backcross progeny in the two F_1 hybrids.

All three species used in the present study are normal homozygotes while the two F_1 hybrids (H_1 and H_2) are structural heterozygotes as evidenced by the presence of the association of four or three chromosomes in these PMCs. The reduced chiasma frequencies in the two hybrids indicates reduced homologies between the corresponding parental taxa.

The occasional occurrence of twelve bivalents in a PMC in the two F_1 hybrids suggests that despite morphological divergence, the species have retained sufficient ancestral homologies. The low frequency of the higher chromosome associations, and inversions, when present, suggests that the altered segments are not large. Univalent formation in H_2 may be due to the failure of pairing between the chromosomes of the two corresponding parents, which are structurally different. Alternately, these may have resulted from desynapsis of some or all of the chromosomes forming multivalents and bivalents at pachytene and diakinesis. Desynapsis leading to univalent formation may be one of the isolating mechanisms between the *C. annuum* and *C. baccatum* of the present study.

Desynapsis as a isolating mechanism has been reported earlier in the interspecific hybrids of *Oryza* (Shastri et al.

1964). A single persistent bridge with or without fragments was observed in H_2 , which is suggestive of inversion heterozygosity.

A variation in pollen fertility in F_1 hybrids (H_1 and H_2) reflects the differences in the cytogenetic architecture of the concerned parents. It is possible that the parents involved in hybridization differed with respect to several genes, a disharmonious assemblage of which might have largely influenced pollen fertility.

Pickersgill (1977) obtained a partially sterile F_1 hybrid between cultivars of *C. annuum* and *C. chinense*. Bivalents and univalents alone were recorded in the F_1 hybrid. She attributed the sterility of the F_1 hybrid to small scale rearrangements of the chromosomes of the two species and also to the cultivated forms. Koompai (1976) and Tanksley (1984) also reported partially sterile F_1 hybrids between *C. annuum* and *C. chinense*. They believed that the two species differ from each other by a single translocation and some minor structural alterations. Egawa and Tanaka (1986) reported a sterile F_1 hybrid between *C. annuum* var. 'minimum' and *C. baccatum* var. 'baccatum'. They concluded that genomes of the two species differed from each other by at least three translocations.

The present cytogenetic analysis of the two F_1 hybrids (H_1 and H_2) indicates that *C. annuum* differs from *C. chinense* by two translocations and minor structural alterations and from *C. baccatum* var. 'pendulum' by two translocations, one inversion and some minor structural alterations.

The differences recorded in the two F_1 hybrids between the present and earlier studies of Pickersgill (1977), Koompai (1976), Tanksley (1984) and Egawa and Tanaka (1986) can be attributed to the inherent genomic differences in the respective taxa employed in the hybridization programme. The results suggest that chromosomal differentiation has occurred between varieties of one or both of the respective species employed in the cross. The inability to obtain reciprocal hybrids in the present investigation can be attributed to reasons other than incompatibility. It is possible that the cytoplasm of the ovules of *C. chinense* or *C. baccatum* may react with gene/genes of *C. annuum* pollen, hindering a viable hybrid.

Meiotic irregularities and partial sterility encountered in the two F_1 hybrids (H_1 and H_2) may be attributed to structural differences found in the respective parents. Such structural changes can inhibit the production of fertile hybrids, as in the present case, and may provide the necessary isolation for the development of diverse species by a further accumulation of genetic differences.

Finally, these structural alterations found in the H_1 and the desynapsis found in the *C. annuum* × *C. baccatum* hybrid acted as effective isolation barriers, in addition to the other isolation mechanisms mentioned earlier, in preventing natural hybridization among the species of the present study. This suggestion gains support from the fact that no naturally occurring hybrids have been reported when these species were grown sympatrically (Pickersgill 1971).

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References

- Aniel Kumar O (1984) Cytogenetical and Palynological studies in the genus *Capsicum* L. Ph D Thesis, Andhra University, Waltair
- Anonymous (1983) Genetic resources of *Capsicum*. Int Board Plant Genet Res, Rome, Italy
- Egawa Y, Tanaka M (1984) Cytogenetic relationship among three species of chili peppers, *Capsicum chinense*, *C. frutescens* and *C. baccatum*. Jpn J Breed 34:50–56
- Egawa Y, Tanaka M (1986) Cytogenetical study of the interspecific hybrid between *Capsicum annuum* and *C. baccatum*. Jpn J Breed 36:16–21
- Eshbaugh LH (1964) A numerical taxonomic and cytogenetic study of certain species of the genus *Capsicum*. Ph D Thesis, Indiana University
- Eshbaugh LH (1975) Genetic and biochemical systematic studies of chili peppers (*Capsicum*-Solanaceae). Bull Torrey Bot Club 102:396–403
- Eshbaugh LH (1980) The taxonomy of the genus *Capsicum* (Solanaceae). Phytologia 47:153–166
- Hirose T, Nishi S, Takashima S (1960) Studies on the interspecific crossing in cultivated *Capsicum*. I. Crossability. Sci Rep Kyoto Prefect Univ Agric 12:40–46
- Koompai P (1976) Some barriers to interspecific crossing and gene exchange in five species of *Capsicum*. MSc Thesis, Reading University
- Lippert LF, Smith PG, Bergh BO (1966) Cytogenetics of the vegetable crops. Garden pepper *Capsicum* sps. Bot Rev 32:24–55
- Ohta Y (1961) Cytogenetical studies in the genus *Capsicum*. I. *C. frutescens* × *C. annuum*. Jpn J Genet 36:105–111
- Panda RC (1985) Cytogenetic studies in chili pepper (*Capsicum* L.). Ph D Thesis, Andhra University, Waltair
- Pickersgill B (1971) Relationships between weedy and cultivated forms in some species of chili peppers (genus *Capsicum*). Evolution 25:683–691
- Pickersgill B (1977) Chromosomes and evolutions in *Capsicum*. In: Pochard E (ed) 'Capsicum 77' C R 2me Congr Eucarpia Piment, Avignon-Montfavot, France, pp 27–37
- Shastry SVS (1964) Chromosome structural differentiation, isolating mechanism and speciation in *Oryza*. In: Rice genetics and Cytogenetics. Elsevier, Amsterdam London New York, pp 111–117
- Smith PG, Heiser Jr CB (1951) Taxonomic and genetic studies on the cultivated peppers, *Capsicum annuum* L. and *C. frutescens* L. Am J Bot 38:362–368
- Smith PG, Heiser Jr CB (1957) Breeding behaviour of cultivated peppers. Proc Am Soc Hortic Sci 70:286–290
- Somanatha Pillai ER, George MK, Mercy ST (1977) Studies on interspecific hybrids of five species of *Capsicum* with special reference to its qualitative and quantitative characters. Agric Res J Kerala 15:1–5
- Tanksley SD (1984) Linkage relationships and chromosomal locations of enzyme-coding genes in pepper *Capsicum annuum*. Chromosoma 89:352–360
- Venkateswara Rao K, Yesoda Raj A (1974) Interspecific crosses in genus *Capsicum*. J Res APAU 11:1–10
- Yamamoto N (1978) The origin and domestication of *Capsicum*. Ph D Thesis, Kyoto University